

Yeager
1985

RICHARD I. NEWSOM

Fish hosts

STENOTOPIC HOST REQUIREMENTS FOR GLOCHIDIA OF
EPIOBLASMA (=DYSONOMIA = PLAGIOLA) CAPSAEFORMIS,
E. INTERRUPTA AND E. TRIQUIETRA (PELECYPODA: UNIONIDAE)

in the Powell River, upper Tennessee River drainage

Bruce L. Yeager
Field Operations, Eastern Area
Division of Services and Field Operations
Office of Natural Resources and Economic Development
Norris, Tennessee

This needs more work

who uses E. interrupta?

May 1985

ABSTRACT

three freshwater mussel species,

In May and June of 1984, gravid females of Epioblasma capsaeformis,
E. interrupta and E. triquetra were collected from the Powell River, in
Tennessee. Laboratory Induced infestations ^{with} of the parasitic ^{of} glochidia
stage of these mussels ^{on fishes} resulted in the identification of the
following suitable hosts for Epioblasma capsaeformis; Etheostoma maculatum,
E. rufilineatum, Percina sciera and Cottus carolinae, ~~six~~ species of fish:
Etheostoma blennioides, E. maculatum, E. rufilineatum, E. simoterum,
Percina caprodes and C. carolinae, yielded juveniles of Epioblasma interrupta,
in infestation trials. Juveniles of Epioblasma triquetra transformed
from P. caprodes and C. carolinae. ^{The apparent} stenotopic requirements ^{of riffle-dwelling} for perciform
hosts ^{for Epioblasma} inhabiting riffles ^{account for} may be contributory to the historical decline of
the genus Epioblasma.

in North America

and the loss of riffle habitat through river modifications

Re-write badly needed

more names + verified

The freshwater mussel genus Epioblasma ^{includes} ~~is comprised of~~ seventeen species, all but one of which occurred historically in the Tennessee River system. The pattern of distribution for the genus indicates that all members are of Tennessee-Cumberland river origin, either belonging to the ancient Cumberlandian fauna (Ortman 1924, 1925) ~~or being of descent from~~ ^{of descending flow} ~~it~~ (Johnson 1978). Species of Epioblasma are characteristic ^{typical} inhabitants of riffles ^{with} ~~in areas of~~ swift current in medium and large rivers. The majority of species in Epioblasma are either presumed extinct (Johnson 1978) or ~~are~~ ^{anthropogenic} endangered from ~~activities of man such as pollution and construction of impoundments, which impact or destroy these shoal~~ ^{ostensibly that after} habitats ⁱⁿ ~~of~~ larger rivers. All species of mussels extirpated from the Ohio River drainage (Stansberry 1970) ^{belonged to this genus} ~~are members of Epioblasma.~~

Of the surviving members of the genus, E. capsaeformis and E. interrupta are restricted in distribution to the Tennessee and Cumberland River systems. Both species are absent from the mainstream Tennessee River, rare in tributaries to the Cumberland River, and locally ^{rare} common ~~or abundant~~ in ^{selected} the tributaries of the Tennessee River. Occurring in the upper White, Missouri, Mississippi, Illinois, Tennessee, Cumberland, Ohio, Green and St. Lawrence river systems and Lakes Michigan and Erie. E. triquetra is the most widely distributed and abundant member of the genus,

Incidental to life history studies on the endangered dromedary mussel (Dromas dromas) and the Cumberland monkeyface (Quadrula intermedia) in the Powell River of Tennessee, gravid females of ~~the three species~~, E. capsaeformis, E. interrupta and E. triquetra were collected by biologists of the Tennessee Valley Authority. Knowledge of the reproductive biology of species from the Cumberlandian mussel fauna (Zales and Neves 1982, Yeager and Neves 1985) and particularly Epioblasma is limited. Investigations of

Needs better organization
 Hell digit only 35 spp remaining in TN
 drawings, sections E. f. rubinosculum
 drawings plans of Epioblasma spp, etc

cite any work of Epioblasma biology?
 sexual dimorphism

mussel-fish host relationships in unionid life cycles are imperative for any effective conservation efforts. The objectives of this ^{study} report are to provide information on the period ^{of} when gravid female ^{study} Epioblasma contain mature glochidia; to identify suitable fish hosts for the glochidia; and to provide description of the early life history stages of the mussels.

METHODS

All specimens of Epioblasma were collected by snorklers ⁱⁿ between river miles 106.8 and 117.4 ^{in Tennessee} ~~of~~ the Powell River ^{near the VA-TN border} in the upper Tennessee River drainage. Gravid females were aged by the external growth ring method (Chamberlain 1931; Crowley, 1957). The mussels were opened slightly by hand or with modified O-ring expanders to check for gravidity. Gravid mussels were placed in small mesh cloth bags and transported in insulated coolers containing river water to a ^{TVA} laboratory in Norris, Tennessee. ~~Until utilized,~~ ^M mussels were held in Living Streams (Frigid Units Inc., Toledo OH) with flow-through spring water at a mean temperature of 17.8 C. ^{photo house,}

When possible, fish were collected from mussel-free sites to ^{avoid} ~~reduce~~ incidences of prior infestations ^{with} of glochidia. Prior to testing, fish were also maintained in Living Stream Units with flow-through spring water. Numbers of fish in each trial depended on species availability. Fish were fed frozen brine shrimp before and during infestation trials.

Mature glochidia were obtained by excision ^{up} of the marsupial gills from gravid female mussels and rupturing the ovisacs with a probe. Glochidia were tested for maturity by exposing a subsample to salt crystals and observing for a strong closing response (Zale and Neves 1982).

Experimental fish were anesthetized with tricaine methanesulfonate (MS-222), and the gills and fins ^{were} inspected for ^{attached} ~~current infestations~~ of ~~contaminant~~ glochidia. Fishes so infested were excluded from experimental infestation trials. Individual fish were infested by pipetting several hundred glochidia into the right branchial chamber. Each fish was exposed to glochidia only once. Mixed species assemblages of fish, [?] infected with a species of mussel, were held in Living Stream units for ten days post-infestation. At five and ten days post-infestation, fish were anesthetized and inspected for retention of glochidia. Fish not found to have retained glochidia were preserved in 10 percent formalin and re-examined. Fish retaining glochidia ^{after} ~~at~~ ten days were sequestered by individual species in 38 L aquaria containing spring water filtered through a bag seive of five micron aperture.

Beginning 11 days post-infestation and every other day thereafter, material from the bottoms of aquaria was siphoned through a 35 micron aperture, nylon mesh sieve. Sloughed glochidia and juvenile mussels were examined ^{under} ~~with~~ a stereomicroscope. Juveniles were placed in gridded 9x9x1.5 cm square petri dishes for observation. Intermittantly, [?] specimens were preserved in five percent formalin buffered to pH 7.0 with ammonium hydroxide. The methods of Castanaga and Kraeuter (1981), as modified by Hudson and Isom (1984), were used to culture juveniles. Juvenile ~~cohorts of~~ mussels were placed in separate 2 L Nalgene trays containing about 1.5 L of [?] feed consisting of a mixed culture of several species of single-celled algae and diatoms obtained from a nearby pond. The pondwater had been filtered through a five micron bag sieve. ~~Food~~ ^{provided} supply was changed daily by filtering the media through a sieve of 35 micron aperture, returning any mussels poured off from the culture media

and adding the filtered pondwater. Measurements of glochidia and juveniles were obtained under a stereomicroscope equipped with an ocular micrometer.

RESULTS

Adults of the three species of Epioblasma were usually found lying on the substrate surface or only partially buried. ^{when?} All three species were collected ⁱⁿ from the swifter currents of large riffles in firm, gravel-cobble substrate. Sites and numbers of females of each species collected ^{are summarized} appear in Table 1.

Gravid specimens of E. interrupta and E. triquetra were found from May 1 to June 5, ^{at} in water temperatures ranging from 15.0 to 17.8 C. During this period 58 percent ^{of} E. capsaeformis, 45 percent of E. interrupta and 31 percent of E. triquetra females were gravid. Gravid E. capsaeformis were collected only until May 18. Unlike the two other species, females of E. capsaeformis lay partially open, displaying a brilliant "electric blue" mantle that was often observable by collectors ^{from the stream bank} without the aid of a viewing box or snorkeler's mask.

^{Estimated} ages of the gravid females collected were as follows ^{of} each species: E. capsaeformis, 7 to 10 years; E. interrupta, 8 to 13 years; and E. triquetra, 5 to 10 years. For all three species, only the outer demibranchs served as marsupia. Water tubes containing mature glochidia were swollen, whitish and granular in appearance. Unfertilized eggs accounted for less than one percent of conglomerates.

Fully mature glochidia (Fig. 1 a-c) were nearly transparent, and were subcircular, with a truncated dorsal hinge line. ~~A single adductor muscle was visible.~~ Glochidia are apparently released individually as conglomerates disintegrate, since intact conglomerates could not be teased

from water tubes. Only glochidia exhibiting a strong, immediate closing response when exposed to salt crystals were deemed satisfactory for infestation trials. Mean dimensions of 100 mature glochidia from two females of each species were: E. capsaeformis, length 0.25 mm (SE=.06), depth (dorsal to ventral) 0.23 mm (SE=.05), hinge length 0.17 mm (SE=.05); E. interrupta, length 0.24 (SE=.01), depth 0.23 (SE=.05), hinge length 0.17 (SE=.04); E. triquetra, length 0.22 mm (SE=.03), depth 0.22 mm (SE=.02), hinge length 0.16 mm (SE=.03).

any observable differences in size of skin

Glochidia typically attached to the distal portion of gill lamellae on experimental fish. ^{at mouth} An exception was that on specimens of the banded sculpin, Cottus carolinus, most glochidia attached and encysted in the epithelial tissue lining the branchial cavity. Degrees of infestation were light to moderate depending on relative size or species of fish. Immediately after infestation, as few as six to ten glochidia were observed on some species of percids or cyprinids and as many as several hundred on centrachids or larger percids.

Eighteen species of fish (Table 2) were exposed to glochidia of E. capsaeformis in laboratory trials. Thirteen species of fish sloughed all glochidia by ten days post-infestation. Of the remaining five species, Etheostoma maculatum, E. rufilineatum, Percina sciera and Cottus carolinus were identified as hosts for glochidia of E. capsaeformis. Periods for transformation (Table 2) varied from nineteen to thirty-four days. The Tennessee snubnose darter, Etheostoma simoterum retained glochidia for up to fourteen days but produced only unviable, partially transformed glochidia.

five common names also

Twenty-six species of fish (Table 3) were exposed to glochidia of E. interrupta. By 10 days post-infestation, twenty species of fish sloughed

Juveniles were obtained
all glochidia. ~~Mussels transformed from~~ six species of fish: Etheostoma
blennioides, E. maculatum, E. rufilineatum, E. simoterum, Percina caprodes
and Cottus carolinae. Transformation occurred in 16 to 45 days. Juveniles
from the log perch, P. caprodes were initially more vigorous and exhibited
a greater amount of foot activity and movement than juveniles transformed
on other species of fish. Of the 25 fish (Table 4) exposed to glochidia
of E. triquetra, six species retained glochidia for ten days. Only two
suitable hosts, P. caprodes, the log perch, and Cottus carolinae, the banded
sculpin, were identified. Juveniles, equally vigorous from both these fish
species, transformed in 24 to 44 days.

In size and shape, newly metamorphosed juveniles (Fig. 2 a,b,d) of
Epioblasma spp. were nearly identical to glochidia. However two adductor
muscles, gill lamellae, functioning cilia, and other internal organs were
apparent. Juveniles moved by extending the extremely adhesive foot, then
drawing the body to the foot. By one to three days after dropping off host
fish (Fig. 2c) juveniles showed evidence of shell growth. Twenty-seven day
old juveniles of E. interrupta had the characteristic "hatchet" shaped foot
of adult mussels. Culturing of E. capsaeformis and E. interrupta ~~was~~
ended at 30 days. When all culturing activities were terminated at 67 days (Fig.
2e), juveniles of E. triquetra were surviving in the food culture and con-
tinued to show growth.

Results don't read smoothly

DISCUSSION

Field investigations ~~from~~^{from} the Powell River support the status
of Epioblasma as a genus of long-term or bradytictic breeders (Ortman 1919).
~~In our study~~ [?] All three species of Epioblasma were gravid with mature glochidia

in May or June and were nongravid by mid-July. ^{spent} Seven females of E. triquetra reported from the Ohio River in September and October were gravid (Ortman 1912). In the Powell River, females of E. capsaeformis ceased displaying the "electric blue" mantle, a feature not shown by the other two species; buried deeper; and were nongravid sooner, indicating a cessation of spawning earlier than either E. interrupta or E. triquetra. } ambward

Laboratory-induced infestations ^{with} of glochidia ~~with~~ ^{of} the three species of Epioblasma indicated a relatively high degree of host specificity, restricted to ~~a narrow range of~~ ^{relatively few} perciform fishes. Within the family Percidae encompassing hosts, three congeneric, sympatric species of Percina and two of Etheostoma produced no juveniles in infestation trials of Epioblasma capsaeformis. None of the six species of Etheostoma and a congeneric Percina ~~also of the family Percidae~~ yielded juveniles of Epioblasma triquetra. } ambward

Transforming on six perciform species, Epioblasma interrupta was only slightly less fastidious in required host species than the other two mussel species. Two darters congeneric with demonstrated hosts of E. interrupta were unsuitable as hosts. [Species of Percina (3) and Etheostoma (2) congeneric with demonstrated hosts of E. capsaeformis produced no juveniles in laboratory tests.] Several percid species, e.g. Etheostoma jessiae, E. caeruleum and E. simoterum however did retain glochidia of Epioblasma capsaeformis and triquetra for longer periods than other nonhost fish. As suggested for other lampsiline mussels (Zale and Neves, 1982) retention of glochidia by these nonhost fish species ^{may} be related to biochemical or physiological similarities ^{among closely related} within taxa containing hosts.

Explanations for the range ^{of observed} fish host specificity required by mussel species are at present speculative. Whatever the etiology, the degree of host specificity may affect the chances of survival for mussel ^{es}

species in the face of largescale alterations of aquatic habitats by man. Anodonta grandis, a characteristic, successful member of the subfamily Anodontinae, has a broad tolerance (Parmalee 1967, Clark 1973, Mathiak 1979) of both lentic and lotic habitats and exhibits eurytopic use of fish hosts, exceeding 30 species in number (Fuller 1974, Trdn and Hoeh 1982). Neves et al. (1985) recognized that mussels of the subfamily Amblyminae have a less generalist life history, i.e. glochidia exhibit a greater degree of host specificity and are more habitat specific in requirements. Fish hosts ~~of~~ mussel species in this subfamily are fewer in number and are more closely related phylogenetically. Zale and Neves (1982) reported two species of Villosa, a genus of amblyminae, to parasitize only 3 of 24 fish species in their area of study. Narrow spectrum host requirements were also reported by these authors for Medionidus conradicus and Lampsilis fasciola. Similarly, stenotopic use of hosts was found for the three species of Epioblasma, herein reported.

A general decline of unionid mussel species in the last century has been well documented (Stansberry 1970, van der Schalie 1975). Many species of the subfamily Amblyminae, particularly those of the unique Cumberlandian fauna, including Epioblasma, have declined in abundance and distribution. Undoubtedly direct destruction and alteration of mussel habitats are major contributors (Stansberry 1970) to declines in ^{now} ~~commercially valuable~~, riffle dwelling populations of mussels.

However, success or decline of various taxa within the Unionidae has also been influenced by the varied reproductive strategies of mussels. Narrow spectrum host requirements by the mussel and use of only those fish hosts occupying riffles, both factors limiting diversity of occurrence and dispersal of juveniles make populations of Epioblasma even more vulnerable

who says Villosa is an amblyminae?
It is Lampsilinae

amblyminae
who says Epioblasma
is Amblyminae

what?
are
fish?

?

to disturbance. The close stenotopic relationship between the species of Epioblasma and their fish hosts was advantageous and successful in undisturbed environments as evidenced by historical records. But decline or loss of even a single host ~~species~~ fish may severely reduce or eliminate recruitment for some species of Epioblasma. As more riffle habitat has been altered by activities of man, the high degree of host specificity exhibited by the three Epioblasma, investigated may have served at least an ancillary role in the historical decline of the genus in general.

ambival

Discussion needs
major rewrite

If ~~improves~~ all over, but questionable for taxonomic genus,
and doesn't make a strong case

ACKNOWLEDGEMENTS

Bennie Kerley and Charles Saylor aided in field collection of gravid specimens. Support for this research was provided by the Tennessee Valley Authority.

LITERATURE CITED

- Castanaga, M. and J. Kraeuter. 1981. Manual for growing the hard clam Mercenaria. Virginia Institute of Marine Sciences. Gloucester Point, VA. 110 pages.
- Chamberlain, T. K. 1931. Annual growth of freshwater mussels. Bull. U.S. Bur. Fish. 46:713-739.
- Clark, A. H. 1973. The freshwater molluscs of the Canadian Interior Basin. Malacologia 13:1-509.
- Crowley, T. E. 1957. Age determination in Anodonta. J. Conchol. 24:201-207.
- Fuller, S. L. H. 1974. Clams and mussels (Mollusca:Bivalvia), p. 215-273. IN: C. W. Hart Jr., and S. L. H. Fuller (eds.). Pollution ecology of freshwater invertebrates. Academic Press, New York.
- Hudson, R. G. and B. G. Isom. 1984. Pearing juvenile mussels of the freshwater mussels (Unionidae) in a laboratory setting. The Nautilus 98(4):129-135.
- Johnson, R. I. 1978. Systematics and zoogeography of Plagiola (= Dysonomia = Epioblasma), an almost extinct genus of freshwater mussels (Bivalvia: Unionidae) from Middle North America. Bull. Mus. of Comp. Zool. Vol. 148 (6):239-321.
- Mathiak, H. A. 1979. A river survey of the unionid mussels of Wisconsin. 1973-1977. Sand Shell Press, Horicon, Wis. 75 p.
- Neves, R. J., L. R. Weaver and A. V. Zale. 1985. An evaluation of host fish suitability for glochidia of Villosa vanuxemi and V. nebulosa (Pelecypoda:Unionidae). Am. Mid. Nat. 113(1):13-19.
- Ortman, A. E. 1912. Notes upon the families and genera of the naiades. Ann. Carnegie Mus. Vol. 8(2):222-365.
- Ortman, A. E. 1919. A monograph of the naiades of Pennsylvania. Pt. 3. Systematic account of the genera and species. Mem. Carnegie Mus. 8(1):1-389, pls. 1-21.
- Ortman, A. E. 1924. The naiad-fauna of Duck River in Tennessee. Amer. Mid. Nat. 9(1):18-62.
- Ortman, A. E. 1925. The naiad fauna of the Tennessee River system below Walden Gorge. Amer. Mid. Nat. 9(8):321-372.
- Parmalee, P. W. 1967. The freshwater mussels of Illinois. Ill. State Mus. Popular Sci. Ser., 8:1-108.
- Stansbery, D. H. 1970. Eastern Freshwater Mollusks (I). The Mississippi and St. Lawrence Rivers systems. Malacologia ~~1970~~ 10(1):9-22.

Trdan, R. J. and W. R. Hoeh. 1982. Eurytopic host use by two congeneric species of freshwater mussel (Pelecypoda:Unionidae:Anodonta). Am. Midl. Nat. 108(2): 381-388.

van der Schalie, H. 1975. An ecological approach to the rare and endangered species in the Great Lakes region. Mich. Acad. Sci. 18:7-22.)

Yeager, B. L. and R. J. Neves. 1985. Reproductive cycle and fish hosts of the rabbit's foot mussel, Quadrula cylindrica strigillata (Mollusca: Unionidae) in the upper Tennessee River drainage. Am. Midl. Nat. (in press).

Zale, A. V. and R. J. Neves. 1982. Fish hosts of four species of lampsiline mussels (Mollusca:Unionidae) in Big Moccasin Creek, Virginia. Can. J. Zool. 60:2535-2542.

maybe
maybe not ?

Table 1. Collections of ~~female~~ ^{females} Epioblasma from the Powell River in 1984. Numbers gravid in parentheses.

<u>Date</u>	<u>Site</u>	<u>Water Temperature °C</u>	<u>Number Females Collected (Number Gravid)</u>		
			<u>E. capsaeformis</u>	<u>E. interrupta</u>	<u>E. triquetra</u>
May 1-2	Bales Ford Island PRM 111.8	15	4 (3)	6 (4)	8 (3)
May 17-18	Fletcher Ford PRM 117.5	16.1	7 (4)	2 (0)	3 (1)
May 19	McDowell Ford PRM 106.5	16.1	1 (0)	-	3 (0)
June 5	Bales Ford Island PRM 111.8	17.8	-	3 (1)	2 (1)
July 19	Bales Ford Island PRM 111.8	23.3	1 (0)	4 (0)	4 (0)
Total Number Inspected			13	15	20

Table 2. Results of laboratory infestations with glochidia of Epioblasma capsaeformis.

Fish Species	No. of Fish Infested	Glochidia Attached		Total Number Juveniles Produced	Period For Transformation (Days)	Mean Holding Temperature °C
		5 Days	10 Days			
<u>Camptostoma anomalum</u>	2	-	-			
<u>Hybopsis amblops</u>	4	-	-			
<u>H. dissimilis</u>	2	-	-			
<u>Notropis coccogenis</u>	1	-	-			
<u>N. galacturus</u>	3	-	-			
<u>N. rubellus</u>	1	-	-			
<u>N. spilopterus</u>	1	-	-			
<u>Pimephales notatus</u>	2	-	-			
<u>Hypentelium nigricans</u>	2	-	-			
<u>Etheostoma jessiae</u>	3	-	-			
<u>E. maculatum</u>	1	x	x	10	19-31	16.2
<u>E. rufilineatum</u>	5	x	x	1	25	15.8
<u>E. simoterum</u>	1	x	x			
<u>Percina caprodes</u>	1	x	-			
<u>P. copelandi</u>	1	x	-			
<u>P. evides</u>	4	x	-			
<u>P. sciera</u>	1	x	x	4	25	15.8
<u>Cottus caroliniae</u>	1	x	x	14	20-34	16.9

Table 3. Results of laboratory infestations with glochidia of Epioblasma interrupta (F. brevidens).

Finally good with YLLS

Fish Species	No. of Fish Infested	Glochidia Attached		Total Number Juveniles Produced	Period For Transformation (Days)	Mean Holding Temperature °C
		5 Days	10 Days			
<u>Camptostoma anomalum</u>	1	-	-			
<u>Hybopsis amblops</u>	1	-	-			
<u>H. dissimilis</u>	2	-	-			
<u>Nocomis micropogon</u>	1	-	-			
<u>Notropis ariomus</u>	3	-	-			
<u>N. chrysocephalus</u>	1	-	-			
<u>N. coccogenis</u>	3	-	-			
<u>N. galacturus</u>	2	-	-			
<u>N. leuciodus</u>	2	-	-			
<u>N. rubellus</u>	2	-	-			
<u>N. spilopterus</u>	2	-	-			
<u>N. telescopus</u>	1	-	-			
<u>Phenacobius uranops</u>	1	-	-			
<u>Pimephales notatus</u>	4	-	-			
<u>Hypentelium nigricans</u>	2	-	-			
<u>Moxostoma erythrurum</u>	1	-	-			
<u>Lepomis auritus</u>	2	-	-			
<u>L. macrochirus</u>	2	x	-			
<u>Etheostoma blennioides</u>	2	x	x	9	34-37	17.0
<u>E. jessiae</u>	3	x	-			
<u>E. maculatum</u>	2	x	x	2	17	15.4
<u>E. rufilineatum</u>	12	x	x	28	16-33	16.3
<u>E. simoterum</u>	10	x	x	3	25-34	16.9
<u>Percina caprodes</u>	1	x	x	104	28-45	16.7
<u>P. evides</u>	3	-	-			
<u>Cottus caroliniae</u>	13	x	x	123	20-48	17.2

Table 4. Results of laboratory infestations with glochidia of Epioblasma triquetra.

Fish Species	No. of Fish Infested	Glochidia Attached		Total Number Juveniles Produced	Period of Transformation (Days)	Mean Holding Temperature °C
		5 Days	10 Days			
<u>Campostoma anomalum</u>	4	-	-			
<u>Hybopsis amblops</u>	1	-	-			
<u>H. dissimilis</u>	2	-	-			
<u>Nocomis micropogon</u>	1	-	-			
<u>Notropis ariomus</u>	1	-	-			
<u>N. coccogenis</u>	2	-	-			
<u>N. chrysocephalus</u>	1	-	-			
<u>N. galacturus</u>	1	-	-			
<u>N. leuciodus</u>	2	-	-			
<u>N. rubellus</u>	2	-	-			
<u>N. spilopterus</u>	2	-	-			
<u>N. telescopus</u>	1	-	-			
<u>Pimephales notatus</u>	2	-	-			
<u>Hypentelium nigricans</u>	3	-	-			
<u>Lepomis auritus</u>	2	x	-			
<u>L. macrochirus</u>	3	x	-			
<u>Etheostoma blennioides</u>	2	x	x			
<u>E. caeruleum</u>	2	x	x			
<u>E. jessiae</u>	1	-	-			
<u>E. maculatum</u>	1	x	-			
<u>E. rufilineatum</u>	9	x	x			
<u>E. simoterum</u>	15	x	x			
<u>Percina caprodes</u>	3	x	x	804	24-33	17.1
<u>P. copelandi</u>	3	x	-			
<u>Cottus caroliniae</u>	4	x	x	26	25-44	17.2

LIST OF FIGURES

- Figure 1. Glochidia of: a. E. capsaeformis
b. E. interrupta
c. E. triquetra

Bar = _____ micrometers

- Figure 2. Juveniles of: a. E. capsaeformis - days
b. E. interrupta - days
c. E. interrupta - days
d. E. triquetra - days
e. E. triquetra - days

Bar = _____ micrometers.